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INTERNATIONAL PRELIMINARY REPORT ON PATENTABILITY
(Chapter II of the Patent Cooperation Treaty)
(PCT Article 36 and Rule 70)

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Applicant MACQUARIE UNIVERSITY et al		

1. This report is the international preliminary examination report, established by this International Preliminary Examining Authority under Article 35 and transmitted to the applicant according to Article 36.
2. This REPORT consists of a total of 4 sheets, including this cover sheet.
3. This report is also accompanied by ANNEXES, comprising:
 - a. ☒ (sent to the applicant and to the International Bureau) a total of 13 sheets, as follows:
 - ☒ sheets of the description, claims and/or drawings which have been amended and are the basis for this report and/or sheets containing rectifications authorized by this Authority (see Rule 70.16 and Section 607 of the Administrative Instructions).
 - ☐ sheets which supersede earlier sheets, but which this Authority considers contain an amendment that goes beyond the disclosure in the international application as filed, as indicated in item 4 of Box No. I and the Supplemental Box.
 - b. ☐ (sent to the International Bureau only) a total of (indicate type and number of electronic carrier(s)) , containing a sequence listing and/or table related thereto, in electronic form only, as indicated in the Supplemental Box Relating to Sequence Listing (see Section 802 of the Administrative Instructions).
4. This report contains indications relating to the following items:

<input checked="" type="checkbox"/> Box No. I	Basis of the report
<input type="checkbox"/> Box No. II	Priority
<input type="checkbox"/> Box No. III	Non-establishment of opinion with regard to novelty, inventive step and industrial applicability
<input type="checkbox"/> Box No. IV	Lack of unity of invention
<input checked="" type="checkbox"/> Box No. V	Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement
<input type="checkbox"/> Box No. VI	Certain documents cited
<input type="checkbox"/> Box No. VII	Certain defects in the international application
<input checked="" type="checkbox"/> Box No. VIII	Certain observations on the international application

Date of submission of the demand 22 April 2005	Date of completion of this report 05 January 2006 10 JAN 2006
Name and mailing address of the IPEA/AU AUSTRALIAN PATENT OFFICE PO BOX 200, WODEN ACT 2606, AUSTRALIA E-mail address: pct@ipaustalia.gov.au Facsimile No. (02) 6285 3929	Authorized Officer DAVID OLDE Telephone No. (02) 6283 2569

Box No. I **Basis of the report**

1. With regard to the language, this report is based on:

☒ The international application in the language in which it was filed☐ A translation of the international application into
translation furnished for the purposes of:

, which is the language of a

☐ international search (under Rules 12.3(a) and 23.1 (b))☐ publication of the international application (under Rule 12.4(a))☐ international preliminary examination (Rules 55.2(a) and/or 55.3(a))2. With regard to the elements of the international application, this report is based on *(replacement sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to this report)*:☐ the international application as originally filed/furnished☒ the description:

pages as originally filed/furnished

pages* 1-11, 13 received by this Authority on 22 April 2005 with the letter of 22 April 2005

pages* received by this Authority on with the letter of

☒ the claims:

pages as originally filed/furnished

pages* as amended (together with any statement) under Article 19

pages* 12 received by this Authority on 22 April 2005 with the letter of 22 April 2005

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☒ the drawings:

pages 1/1 as originally filed/furnished

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☐ a sequence listing and/or any related table(s) - see Supplemental Box Relating to Sequence Listing.3. ☐ The amendments have resulted in the cancellation of:☐ the description, pages☐ the claims, Nos.☐ the drawings, sheets/figs☐ the sequence listing (*specify*):☐ any table(s) related to the sequence listing (*specify*):4. ☐ This report has been established as if (some of) the amendments annexed to this report and listed below had not been made, since they have been considered to go beyond the disclosure as filed, as indicated in the Supplemental Box (Rule 70.2(c)).☐ the description, pages☐ the claims, Nos.☐ the drawings, sheets/figs☐ the sequence listing (*specify*):☐ any table(s) related to the sequence listing (*specify*):

* If item 4 applies, some or all of those sheets may be marked "superseded."

Box No. V Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

1. Statement

Novelty (N)	Claims 1-11	YES
	Claims -	NO
Inventive step (IS)	Claims 1-11	YES
	Claims -	NO
Industrial applicability (IA)	Claims 1-11	YES
	Claims -	NO

2. Citations and explanations (Rule 70.7)

The invention is directed to an isolated *Ophiostoma floccosum* species mutant having enhanced protein secretion capability compared to a parent strain under similar conditions.

The following citations have been considered for the purposes of this report:

D1: Tadesse, *et al.* 2003. Molecular Genetics Genomics. 269:789-796.

D2: Temple, *et al.* 2000. Mycologia. 92(1):1-9.

D3: Temple *et al.* 1997. Fungal Genetics and Biology. 22:39-53.

Novelty(N) and Inventive Step(IS)

Claims 1-11 are considered novel and inventive in light of documents D1-D3. The citations disclose the characterisation of cerato-ulim (CU) secretion by *Ophiostoma* strains. However the secretion is to the cell surface and not the extracellular medium according to the Attorneys comments during International Examination. The present invention is directed to secretion of proteins to the extracellular medium. Therefore the claimed invention is considered novel and inventive in light of D1-D3.

Industrial Applicability(IA)

Claims 1-11 meet the requirements of the PCT in terms of Industrial Applicability.

Box No. VIII Certain observations on the international application

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:

Claims 1, 2, 5-11 are not fully supported by the description. These claims encompass any strain of *Ophiostoma* that has been mutated by any means having enhanced protein secretion capability. In contrast the description only provides support for specific mutant strains as defined in claims 3 and 4. The applicant has not disclosed or demonstrated a general principle applicable to all strains of *Ophiostoma* as the description only discloses one parent strain from which a number of mutant strains have been selected. Therefore as the claims are not limited to the disclosed mutant strains of *Ophiostoma floccosum* the scope of the claims is not fully supported by the description.

FUNGAL HOSTS FOR EXPRESSION OF RECOMBINANT PRODUCTS

Technical Field

The present invention relates to fungal hosts having enhanced protein secretion.

5 Background Art

The ascomycete *Ophiostoma* spp. comprises common sap-staining fungi such as *O. piceae* that cause discoloration of wood. Some albino variants of *O. floccosum* have been used as biological control agents to prevent sap-staining (Harrington *et al.*, 2001. Mycologia, 93:110-135). A fungus capable of invading wood via the parenchyma cells
10 must secrete enzymes outside the growing mycelium in order to utilize the non-structural components of sapwood (eg. sugars, proteins and extractives). Secreted enzymes such as lipases (Brush *et al.*, Bio. Med. Chem. 7:2131-2138, 1999) and proteinases (Abraham and Breuil, Enzyme Microb. Technol. 18:133-140, 1996) from *Ophiostoma* have been studied in some detail. However, in addition to these studies, very little has been
15 published about biochemical and genetic properties of *Ophiostoma*.

The present inventors have developed *Ophiostoma* mutants as hosts for expression of industrially important enzymes and biomolecules. *Ophiostoma* mutants can be used for new expression systems.

20 Disclosure of Invention

In a first aspect, the present invention provides an isolated mutant *Ophiostoma* species having enhanced protein secretion capability as compared with its parent strain cultured under similar conditions.

Preferably, the isolated *Ophiostoma* species is *Ophiostoma floccosum*. More
25 preferably, the *Ophiostoma floccosum* is selected from mutant strains J2026MQ.1.1, J2026MQ.1.2, J2026MQ.2.1, J2026MQ.3.1, J2026MQ.4.1, J2026MQ.5.1 and J2026MQ.5.5, as herein defined.

Mutants J2026MQ.1.1, J2026MQ.1.2, J2026MQ.5.1 and J2026MQ.5.5 were deposited at the National Measurement Institute (NMI) (formally Australian Government
30 Analytical Laboratory (AGAL)) on 17 September 2004 under accession numbers as set out below.

Mutant designation	NMI Accession Number
J2026MQ.1.1	NM04/42878
J2026MQ.1.2	NM04/42879
J2026MQ.5.1	NM04/42880
J2026MQ.5.5	NM04/42881

In a more preferred embodiment, the isolated *Ophiostoma* species is *Ophiostoma floccosum* strain J2026MQ.1.1, or J2026MQ.1.2 or J2026MQ.5.1 or J2026MQ.5.5 or any of their descendants or mutant thereof.

5 The present inventors have found that mutant strains having enhanced protein secretion capability are particularly suitable for receiving and harbouring expression vectors and producing recombinant products.

 Preferably, the protein is an enzyme. More preferably, the enzyme is selected from protease, amylase, lipase, glucoamylase, β -galactosidase, and β -glucosidase.

10 Preferably, the mutant *Ophiostoma* species according to the present invention is characterised by:

- (a) one nucleus per conidium/blastospore;
 - (b) conidia having mean spore size no less than about 2-3 μ m in diameter; and
 - (c) being capable of secreting at least about two times more of a selected protein
- 15 into culture medium when compared to the secretion by the parent strain grown under similar conditions.

 Preferably, the mutant is capable of secreting at least about three times more of a selected protein when compared to the secretion of the parent strain grown under similar conditions.

20 Preferably, the selected protein is a proteinase.

 The isolated *Ophiostoma* species has preferably undergone modification / selection to alter its ability to carry out or perform protein secretion.

It has been found by the present inventors that modifying / altering DNA by random mutagenesis creates a more adaptable *Ophiostoma* species to act as a host for gene expression.

5 Mutant *Ophiostoma* species according to the present invention differ from naturally occurring *Ophiostoma* species in their enhanced ability to secrete proteins into the cultivation medium.

In a second aspect, the present invention provides use of a mutant fungal species according to the first aspect of the present invention in an industrial process such as pulping, bleaching, recombinant protein production, and any other suitable process
10 utilizing microbial production or action.

An advantage of using a fungal species (*Ophiostoma*) instead of bacteria or yeasts is that *Ophiostoma* is naturally living on wood for *in situ* delivery of selected gene products.

The present invention is particularly suitable for developing modified fungal
15 species which are capable of secreting extracellular components such as enzymes that can be applied to processes either as liquid form from submerged fermentation or delivered *in situ* by the fungus growing on a particular material in solid state.

Any discussion of documents, acts, materials, devices, articles or the like which has been included in the present specification is solely for the purpose of providing a
20 context for the present invention. It is not to be taken as an admission that any or all of these matters form part of the prior art base or were common general knowledge in the field relevant to the present invention as it existed in Australia before the priority date of the claims of the invention.

Throughout this specification, unless the context requires otherwise, the word
25 "comprise", or variations such as "comprises" or "comprising", will be understood to imply the inclusion of a stated element, integer or step, or group of elements, integers or steps, but not the exclusion of any other element, integer or step, or group of elements, integers or steps.

In order that the present invention may be more clearly understood, preferred
30 forms will be described with reference to the following drawings and examples.

Brief Description of the Drawings

Figure 1 shows SDS-PAGE separation of the proteins secreted by *Ophiostoma* J2026 and selected first round mutants. M, molecular weight markers. Lane 1, strain J2026; lanes 2-9, strains J2026MQ.1.1 - J2026MQ.1.8, respectively.

5 Figure 2 shows PCR analysis of *Ophiostoma* colonies following protoplast transformation. L, 1 kb plus DNA ladder, 1, control plasmid containing *dsRed* gene fragment, 2 and 3, represent non-transformant strains J2026MQ.5.1 and J2026MQ.5.5, respectively, lanes 4 to 18, represent transformants which survived the hygromycin B selection. Only transformants 4, 9, 10, 11 and 15 have the 0.7 kb *dsRed* gene fragment
10 amplified.

Mode(s) for Carrying Out the Invention

In order to obtain the present invention high secreting *Ophiostoma* mutants were made and the second step was to develop an effective transformation system for
15 *Ophiostoma* useful for one or more of three purposes: (i) introduction of novel properties in the fungus for biological control; (ii) introduction of novel enzyme activities such as ligninases in the fungus for biopulping purposes, and (iii) development of *Ophiostoma* as a novel system for the expression of a variety of industrially important gene products, either homologous or heterologous (enzymes, biopharmaceuticals, etc).

20 Some basic requirements for a potential expression host strain, before commencing with biolistic transformation, are: (i) preferably one nucleus per blastospore/conidium; (ii) occurrence of (blasto)spores with a minimum length and diameter of 2-3 μm ; and (iii) sensitivity to a potent antibiotic (eg. hygromycin B) at a manageable level (eg. 50 to 200 U/ml). Experiments on *Ophiostoma floccosum* strains
25 J2026MQ.1.1, J2026MQ.1.2, J2026MQ.2.1, J2026MQ.3.1, J2026MQ.4.1, J2026MQ.5.1 and J2026MQ.5.5 display necessary features and therefore have been chosen as the preferred hosts for further development.

MATERIALS AND METHODS

30

Screening for high secretors

Secretion capacity of filamentous fungi can be improved by random mutagenesis and screening for enhanced and favourable characteristics. This procedure involves the

treatment of fungal spores with physical (UV) or chemical mutagens, followed by screening for better secretors using a plate assay.

A 20 ml spore suspension of *Ophiostoma floccosum* (strain J2026) was placed on a 20 cm glass Petri dish at a concentration of 5×10^8 spores/ml and irradiated at 254 nm UV-light for a period of 0 to 8 min. One ml samples were removed at one minute intervals, diluted in 0.9% NaCl, 0.01% Tween-80 solution and plated on Potato Dextrose Agar + 0.1% Triton X100-plates for counting. Colonies from the 5-50% survival level were picked and patched onto minimal agar plates (Penttilä *et al.*, Gene 61: 155-164, 1987) supplemented with soy hydrolysate (1.5% w/v) and insoluble starch (3% w/v) for the screening of starch-degrading enzymes (e.g. amylase, glucoamylase). The presence of clearing halos around the colonies indicates starch degrading activity. Relative size of the hydrolysis halo around a mutant colony gives an indication of enzyme secretion when compared to the nonmutagenized parent. It will be appreciated that other screening means known to the art can also be used.

Colonies showing considerably larger halos when compared to the parent strain were grown in shake cultures in a medium promoting production of a wide variety of hydrolytic enzymes.

The basal liquid medium for general protein production contained 15 g KH_2PO_4 , 5 g $(\text{NH}_4)_2\text{SO}_4$, 10 ml 100 x trace elements (100 mg $\text{FeSO}_4 \times 7\text{H}_2\text{O}$, 20 mg $\text{MnSO}_4 \times \text{H}_2\text{O}$, 20 mg $\text{ZnSO}_4 \times 7\text{H}_2\text{O}$, 40 mg $\text{CoSO}_4 \times 7\text{H}_2\text{O}$ in 200 ml MQ water) and MQ water to a final volume of 1 litre. The pH was adjusted to 6.5. The basal medium was divided into 50 ml portions in 250 ml conical flasks and soybean flour (consisting of 52% protein, 1% fat) was added to each flask to a final concentration of 1.5% (w/v) for induction of e.g. lipase and protease production. The carbon source used for amylase induction was 3% (w/v) filter sterilized potato soluble starch (Sigma Chemicals). Each flask was inoculated with 10^8 spores and cultivated for 5 days at 28°C and 250 rpm.

Enzymes secreted into the cultivation medium were assayed using previously published methods. Protease activity was monitored as in (Lovrien *et al.* J. Appl. Biochem. 7:258-272, 1985) and the method for amylase and glucoamylase activity has been described in Bailey and Nevalainen (Enzyme Microb Technol. 153-157, 1981). SDS-PAGE analysis of secreted proteins from the culture supernatants was carried out in 12.0 % (w/v) and 4-20 % (w/v) gradient acrylamide gels according to Laemmli (Nature 227, 680-685, 1970).

Biolistic and Protoplast transformations

Transformation was carried out using particle bombardment of fungal spores as outlined in Hazell *et al.* (Lett. Appl. Microbiol. 30:282-286, 2000) using a hepta-adaptor device that allows delivery of DNA into the conidia from seven barrels, thereby increasing the transformation efficiency (Te'o *et al.*, J. Microbiol. Methods, 51:393-399, 2002). Seven portions of 7 to 14 days old spores (1×10^7 – 1×10^8 in 0.9 % NaCl, 0.01% Tween 80 solution) were plated appropriately on PDA plates for hygromycin selection or MM plates containing acetamide as the sole nitrogen source for the selection of transformants (Penttilä *et al.*, 1987) to align with the seven barrels of the Hepta Adaptor, and left to dry. Precipitation of circular and linear DNA onto tungsten particles (0.7 μ m mean diameter, Bio-Rad) was carried out as described in Hazell *et al.*, (2000) with some minor modifications. For example, 100 μ l of tungsten beads were used instead of 50 μ l per DNA sample and accordingly, the amount of 2.5 M CaCl_2 and 0.1 M Spermidine were increased to 100 and 40 μ l from 50 and 20 μ l, respectively during the precipitation step. Following precipitation, the mixture of tungsten particles coated with DNA were resuspended in 80 μ l of 100 % Ethanol, of which 10 μ l (x 7) samples were removed and used for bombardment as described below. DNA concentrations of 1000 ng (x 7) were used in the experiments.

For bombardment, 10 μ l of DNA-coated tungsten particles were loaded onto seven macrocarrier disks sitting in the seven slots of the Hepta Adaptor holder and left to dry. A single rupture disk of 1350 *psi* strength was placed on the top to temporarily block the helium gas from entering the seven barrels of the Hepta Adaptor instrument. When dried, the Hepta Adaptor holder containing DNA-coated tungsten beads was placed inside the PDS-1000/He system chamber directly below the Hepta Adaptor device. PDA plates containing conidia for bombardment were placed at a target distance of 3 cm.

Following bombardment, PDA plates were incubated at 28 °C for 4-6 hr before overlaying them with 10 ml of PDA containing hygromycin B to a final concentration of 60 U/ml or 90 μ g/ml of G418.

Transformations using protoplasts were carried out as described in Wang *et al.* Mycol. Res. 103:77-80, 1999, with minor modifications. Briefly, a 100 μ l protoplast solution (total of 1×10^8) was gently mixed with 5-20 μ g of transforming DNA and left on ice for 30 mins before plating out using a 7 mL RM (5.0 % yeast extract, 2.0 % glucose, 0.2 % KH_2PO_4) agar overlay onto RM agar plates containing 60 U/mL of Hygromycin B. Plates were left at 28 °C until colonies emerged and were picked and screened for the presence of the transforming DNA using PCR.

RESULTS

Identification and improvement of *Ophiostoma* strains suitable as expression hosts by SDS-PAGE analysis of secreted proteins

5 SDS-PAGE of secreted proteins was initially applied to recognize good protein
secretors suitable for the development of expression hosts and to identify the most highly
secreted proteins to isolate strong gene promoters driving their synthesis (discussed
above). In order to profile the proteins secreted by *Ophiostoma* strains J2026 and
J2026MQ.1.1 - J2026MQ.1.8, culture supernatants were analysed on a 12% SDS-PAGE
10 gel and stained with Coomassie stain (Figure 1) .

In order to develop the strains further, screening for strains with increased protein
secretion and/or modified enzyme profile after UV mutagenesis was carried out. Plate
assays were used for the screening of mutants, for example, with improved starch-
degrading activity.

15 With strain J2026, nine mutant colonies with potentially increased starch
degrading activity were picked from the screening plates and grown in liquid culture as
described in Materials and Methods. Glucoamylase and protease activity as well as total
secreted protein were assayed from the culture supernatants (Table 1).

Table 1, Glucoamylase (GA) and protease activity of first round UV mutants of *Ophiostoma* J2026.

Strain	GA nkat/ml	Protease Units*	Total secreted protein (mg/ml)
J2026 (parent)	10.3	4.43	0.228
J2026MQ.1.1	36.83	0.25	0.268
J2026MQ.1.2	18.4	13.25	0.236
J2026MQ.1.3	18.23	13.03	0.213
J2026MQ.1.4	13.35	4.97	0.220
J2026MQ.1.5	11.91	7.12	0.287
J2026MQ.1.6	10.48	0.48	0.200
J2026MQ.1.7	9.99	13.41	0.205
J2026MQ.1.8	9.53	5.81	0.246
J2026MQ.1.9	7.41	2.86	0.269

* (as in Lovrien *et al.* 1985)

5 The mutant J2026MQ.1.1 showed about three fold increase in the glucoamylase activity and 18 fold decrease in the protease activity when compared to the nonmutagenised parent J2026. In addition, secretion of the total protein was increased by 14 % in J2026MQ.1.1. In the mutant J2026MQ.1.5, the glucoamylase and protease activities were slightly increased when compared to the parent J2026, however, the total
10 secreted protein was increased by 25 %. These examples show that total protein secretion and protein profiles of *Ophiostoma* can be modified by random mutagenesis. Low protease producing strains are of special interest as expression host for foreign gene products.

15 Figure 1 shows the proteins secreted by the mutants in Table 1, separated by SDS-PAGE highlighting the different protein profiles.

Protease enzyme activity secretion profiles from *Ophiostoma*

20 One important aspect of foreign protein production is proteolytic degradation of the heterologously produced protein by the host proteinases. Having first hand knowledge of the host's proteolytic profile (eg. secreted proteinases) can assist in the

correct decision making in which expression host best suits the gene to be expressed. Selected *O. floccosum* mutant strains plus the parent were cultivated in minimal medium (Penttilä *et al.*, 1987, Gene. 61:155-164) containing starch as a carbon source for 5 days at 28 °C. Proteolytic enzyme activities determined from the culture supernatants are summarized in Table 2. According to the results in Table 2, mutant J2026MQ.1.2 secretes proteins which have strong metallo-like proteinase activity at pH 5, and high serine-like (Chymotrypsin) proteinase activity at pH 8. Culture supernatants were also analyzed on 1D PAGE gels and J2026MQ.1.2 was found to secrete high amounts of a ~30 kDa protein when compared to the parent and to other mutant strains. This 30 kDa protein's N-terminal sequence was determined and found to show similarity to serine-like proteinases from other filamentous fungi. When compared to the parent as well as to J2026MQ.1.2, the mutants J2026MQ.1.1, J2026MQ.2.1, J2026MQ.3.1, J2026MQ.4.1 and J2026MQ.5.1 had higher glucoamylase activities which appeared to be coming from secreted proteins running at molecular weights of 70-80 kDa (also seen in Figure 1). Importantly, the 30 kDa protein (a putative serine proteinase) mentioned above is not present in these high glucoamylase-producing mutants .

Table 2. Summary of secreted proteolytic enzyme activities from selected *Ophiostoma floccosum* strains. J2026 is the parent strain and J2026MQ.1.1, J2026MQ1.2, J2026MQ2.1, J2026MQ3.1, J2026MQ4.1 and J2026MQ5.1 were all mutant strains. Proteinase enzyme activities are represented according to +++++, strong; +++, high; ++, medium; +, low; -+, weak; and --, no activity.

Table 2

Strain	J2026		J2026MQ.1.1		J2026MQ.1.2		J2026MQ.2.1		J2026MQ.3.1		J2026MQ.4.1		J2026MQ.5.1	
	pH 5	pH 8	pH 5	pH 8	pH 5	pH 8	pH 5	pH 8	pH 5	pH 8	pH 5	pH 8	pH 5	pH 8
Protease type														
Metallo	+++	+++	+++	++	++++	++	++	++	++	++	+++	++	+++	++
Serine														
Chymotrypsin	-/+	++	-	+++	+	-	-	-	-	-	-	-	-	-
Elastase	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Subtilisin	-	-	-	-	-	-	-	-	-	-	-	-	-	-
pH	5	7.6	5	7.6	5	7.6	5	7.6	5	7.6	5	7.6	5	7.6
Trypsin	-	-	-	-	-	-	-	-	-	-	-	-	-	-
pH	5	7.5	5	7.5	5	7.5	5	7.5	5	7.5	5	7.5	5	7.5
Cysteine	-	-	-/+	+	-	+++	++	+++	++	+++	++	+++	++	+++

Proteinase enzyme activities are represented according to +++++, strong; +++, high; ++, medium; +, low; -+, weak; and --, no activity.

Transformation of *Ophiostoma* by biolistic bombardment

The average transformation efficiency of *Ophiostoma* was 4 transformants per microgram of DNA using 500 ng of DNA (*hygB* marker), 1350 *psi* and 3 cm target distance. Following bombardment, the transformants were tested on PDA containing 150 units per ml of hygromycin B and 90 µg per ml of G418 or identified by colony growth on minimal medium agar plates containing acetamide as the sole nitrogen source.

Protoplast Transformation of *Ophiostoma*

Using the protoplast approach, approximately 1 in 7.3 colonies had the transforming DNA integrated into the *Ophiostoma* genome (Figure 2). A total of 73 transformants which grew on hygromycin B containing plates were picked and genomic DNA from ten transformants showed to have the ~ 0.7 kb *dsRed* gene fragment integrated into the genomic DNA (Figure 2).

SUMMARY

The present inventors have devised an *Ophiostoma* system for efficient expression of selected gene products. The system comprises promoter(s) functional in *Ophiostoma*, suitable transformation markers and transformation of a suitable *Ophiostoma* strain by biolistic bombardment and by protoplast transformation. The present inventors have shown that the amount of secreted protein of a fungal strain can be increased and protein profiles modified following UV-light mutagenesis. Mutagenesis also allows for the production of host strains with different background enzyme profiles. Enhanced secreting and low protease mutant strain(s) can be used as expression hosts in the production of industrially important molecules or proteins. The transformed mutant strain can be subjected to additional rounds of UV-light mutagenesis to further improve desired characteristics.

It will be appreciated by persons skilled in the art that numerous variations and/or modifications may be made to the invention as shown in the specific embodiments without departing from the spirit or scope of the invention as broadly described. The present embodiments are, therefore, to be considered in all respects as illustrative and not restrictive.

Claims:

1. An isolated mutant *Ophiostoma* species having enhanced protein secretion capability as compared with its parent strain cultured under similar conditions.
2. The mutant according to claim 1 wherein the *Ophiostoma* species is *Ophiostoma*
5 *floccosum*.
3. The mutant according to claim 2 selected from the group consisting of mutant strains J2026MQ.1.1, J2026MQ.1.2, J2026MQ.2.1, J2026MQ.3.1, J2026MQ.4.1, J2026MQ.5.1 and J2026MQ.5.5, as herein defined.
4. The mutant according to claim 2 selected from the group consisting of deposit
10 accession numbers NM04/42878, NM04/42879, NM04/42880, NM04/42881, their progeny, and mutants thereof.
5. The mutant according to any one of claims 1 to 4 capable of receiving and harbouring an expression vector and producing a recombinant product.
6. The mutant according to any one of claims 1 to 5 the protein is an enzyme.
- 15 7. The mutant according to claim 6 wherein the enzyme is selected from the group consisting of protease, amylase, lipase, glucoamylase, β -galactosidase and β -glucosidase.
8. A mutant *Ophiostoma* species is characterised by:
one nucleus per conidium/blastospore;
20 conidia having mean spore size no less than about 2-3 μ m in diameter; and
being capable of secreting at least about two times more of a selected protein into culture medium when compared to the secretion by the parent strain grown under similar conditions.
9. The mutant according to claim 8 capable of secreting at least about three times more
25 of a selected protein when compared to the secretion of the parent strain grown under similar conditions.
10. The mutant according to claim 8 or 9 wherein the selected protein is a proteinase.
11. Use of a mutant fungal species according to any one of claims 1 to 10 in an industrial
30 process selected from the group consisting of pulping, bleaching and recombinant protein production.

Abstract

An isolated mutant *Ophiostoma* species having enhanced protein secretion capability as compared with its parent strain cultured under similar conditions.